

Bioimage Analysis Course 2022

March 7th, 2022

Cell counting

(as an example of image processing workflow)

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“From images of double-stained cells in different conditions, I need to **calculate the percentage of those that are single- or double positive.**”

“I need to **quantify TH-positive neurons** in primary neuronal cultures.”

“I need to separately **quantify the amount of single and double positive cells** in different tissue regions from double labeled immuno images.”

“How to automatically **count stainings** for PV and PNN positive cells?”

“I want to **calculate the number and area** of myocytes in a transversal muscle section.”

“Quantification of **pixel intensity from nuclear envelope vs. cytoplasm** from basic fluorescence microscope”

Cell counting and characterization

Why? (What is your biological question?)

- How many...
 - live / dead cells?
 - cells/droplets/vesicles/bacteria inside a selected area?
- How big...
 - an area/volume?
- How much...
 - protein X is in treated cells vs. control?
 - more protein X is localized in the nuclear envelope relative to cytoplasm?
- How fast...
 - do the cells move/divide/migrate?
- Do the signals colocalize?
 - Colocalization (or co-occurrence) of two different protein clusters

Cell counting and characterization

How? (What are the steps needed to get the result?)

- This lecture: Image processing workflow
- In workshops: How to do it in practice with FIJI ImageJ?

Then what? (What to do with the numbers?)

- Analysis of results, visualizations, statistics
- Conclusions
- Modify protocol, repeat experiment, iterate
- Publish (remember to cite & acknowledge)



Image processing workflow

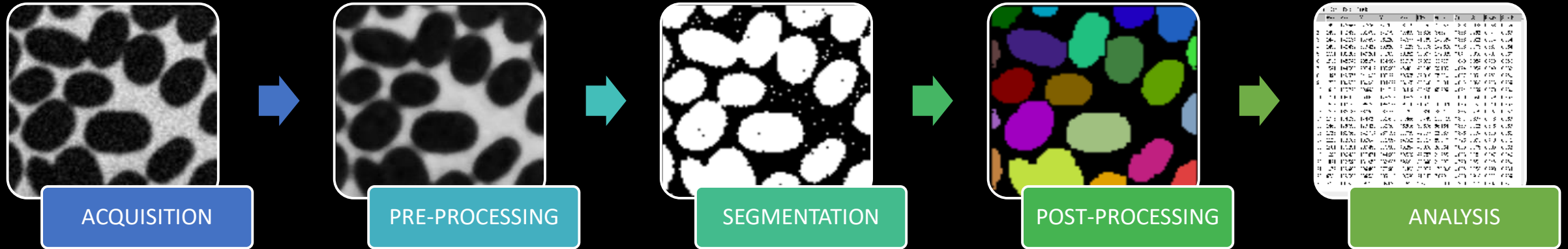
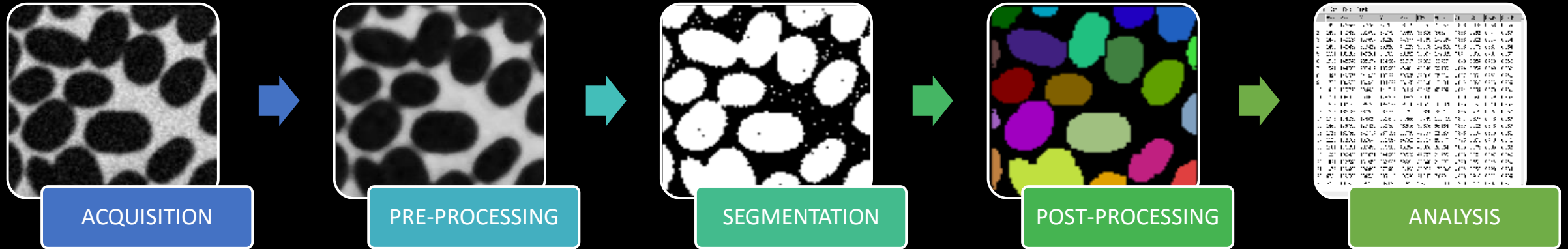


Image processing workflow



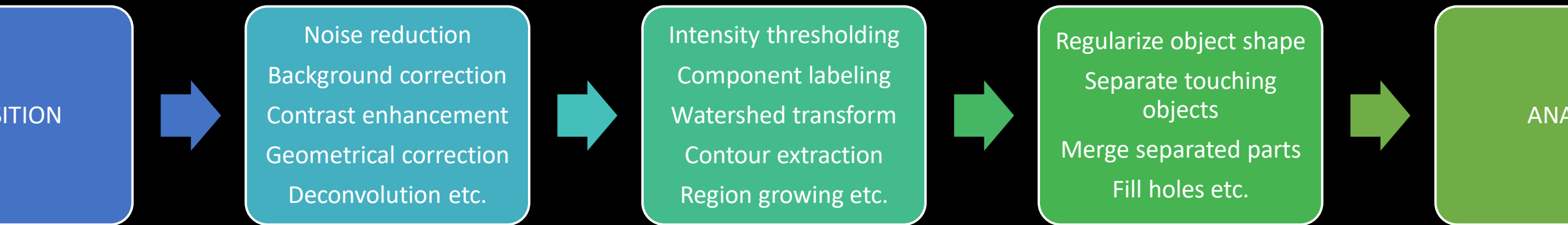
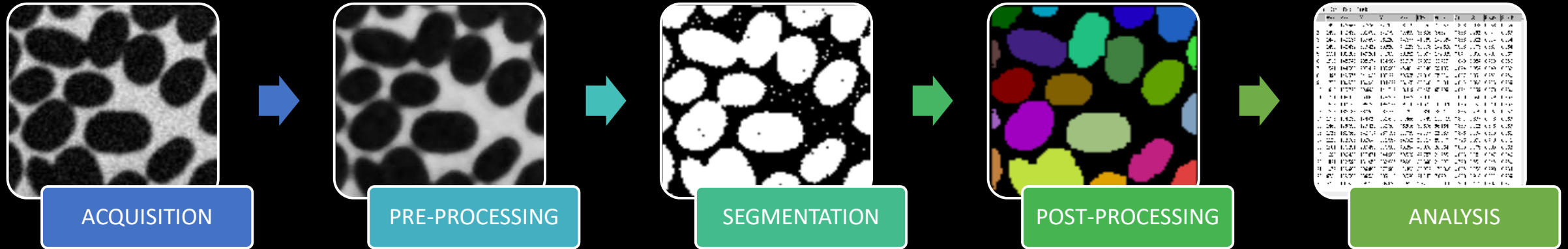
ADJUST PROTOCOL



BIOLOGICAL
QUESTION



Image processing workflow



ACQUISITION



PRE-PROCESSING



SEGMENTATION



POST-PROCESSING



ANALYSIS

Acquisition

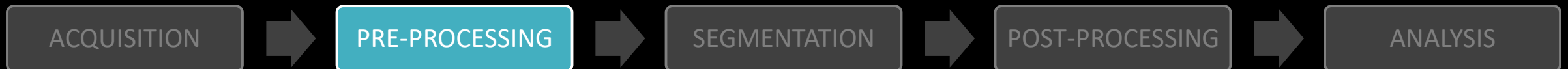
- Very important for good result, but you heard this already...
- Pick the best microscope & acquire optimal images

- "Best" ≠ Newest
- "Best" ≠ Most expensive
- "Best" ≠ Confocal microscope
- "Best" ≠ The one everyone else in the lab is using



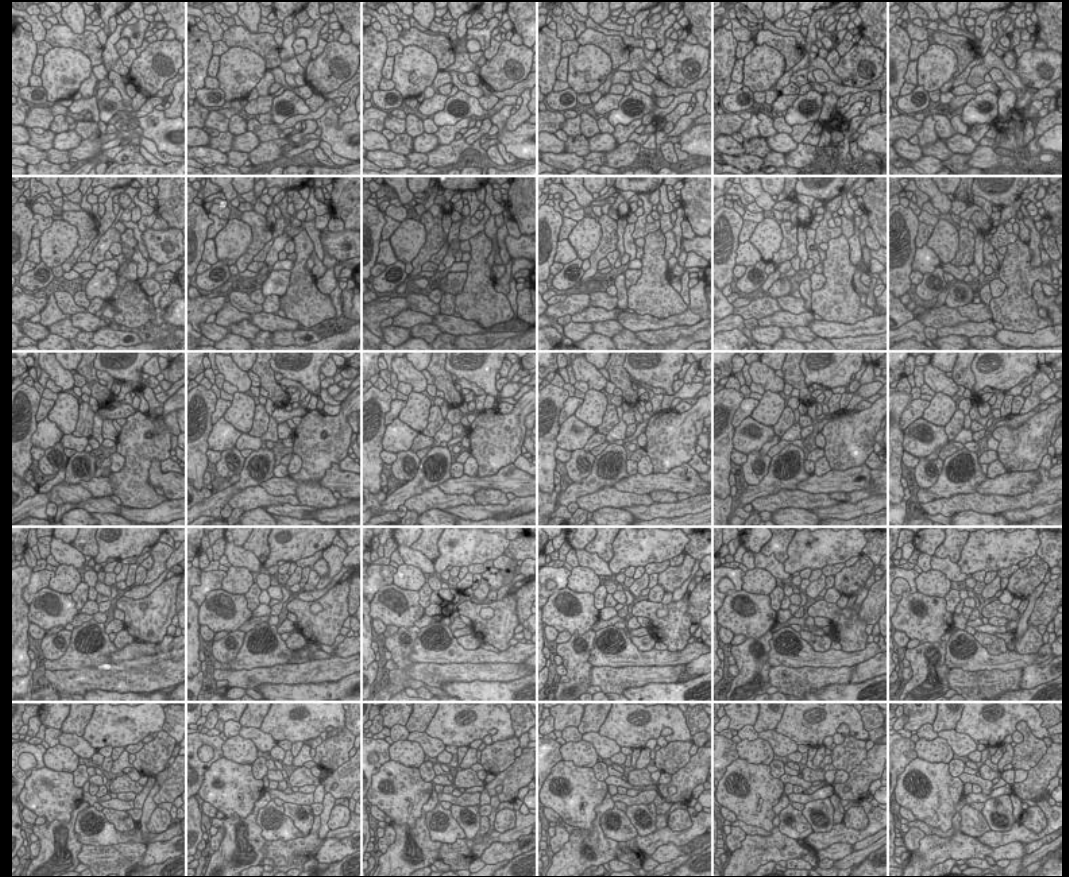
Pre-processing

- Image data format conversion to ensure precise calculations (32 or 64 bit)
- Modify the image using filters to **make later segmentation more effective**
- Which filter(s) to use is highly dependent on your data, but some commonly useful filters include:
 - Normalization
 - Gaussian blur
 - Subtract background
 - Find edges
 - Deconvolution



Pre-processing – Normalization

- Correction of intensity fluctuations during a time series or a z-stack
- In Fiji
 - Plugins → Integral Image Filters → Normalize Local Contrast



ACQUISITION



PRE-PROCESSING



SEGMENTATION



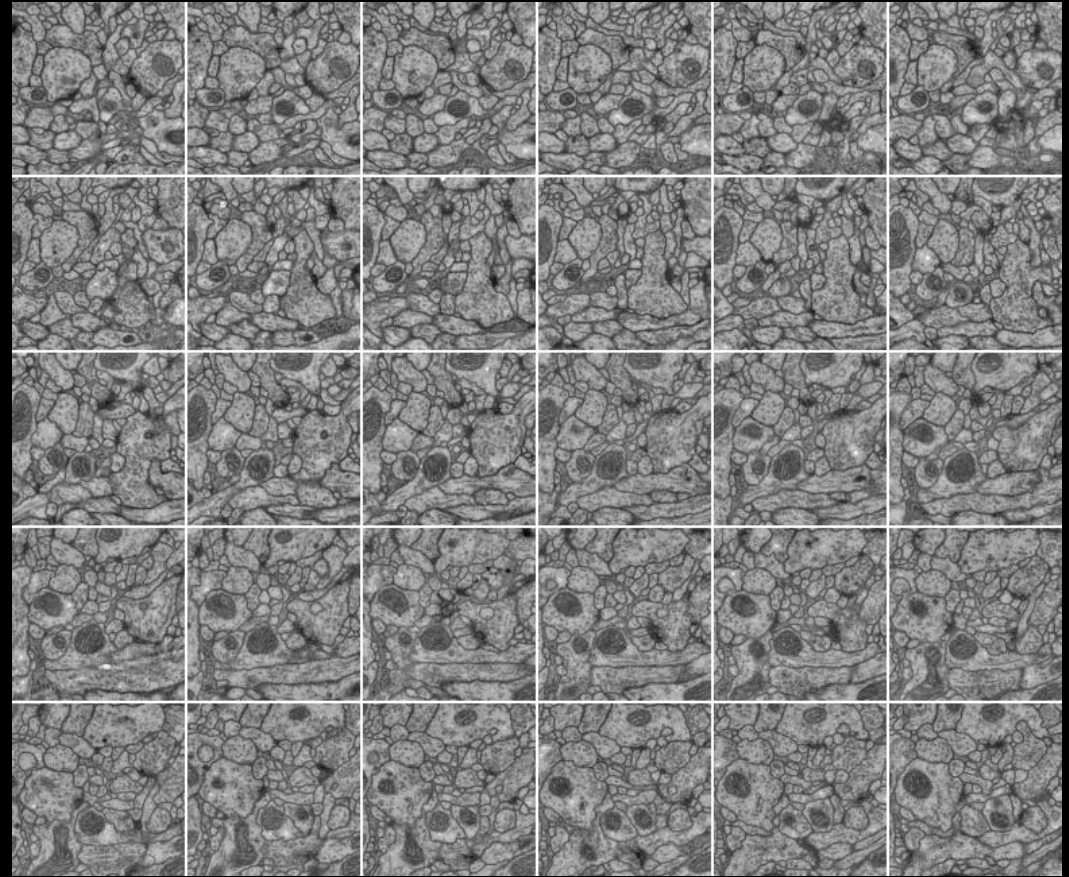
POST-PROCESSING



ANALYSIS

Pre-processing – Normalization

- Correction of intensity fluctuations during a time series or a z-stack
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ACQUISITION



PRE-PROCESSING



SEGMENTATION

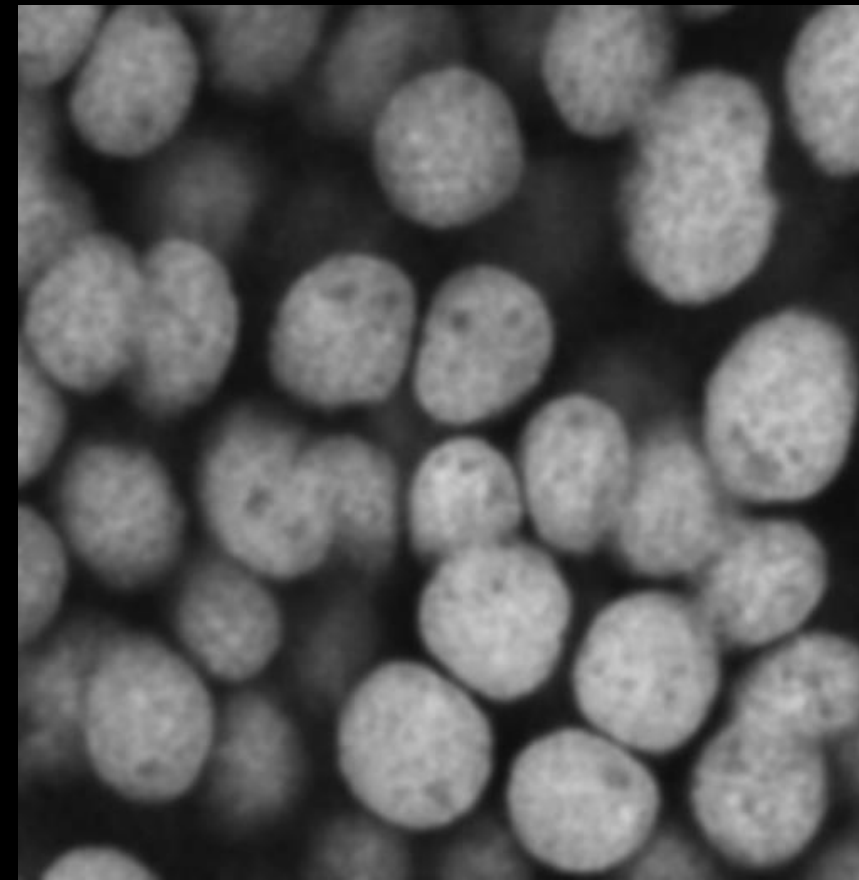
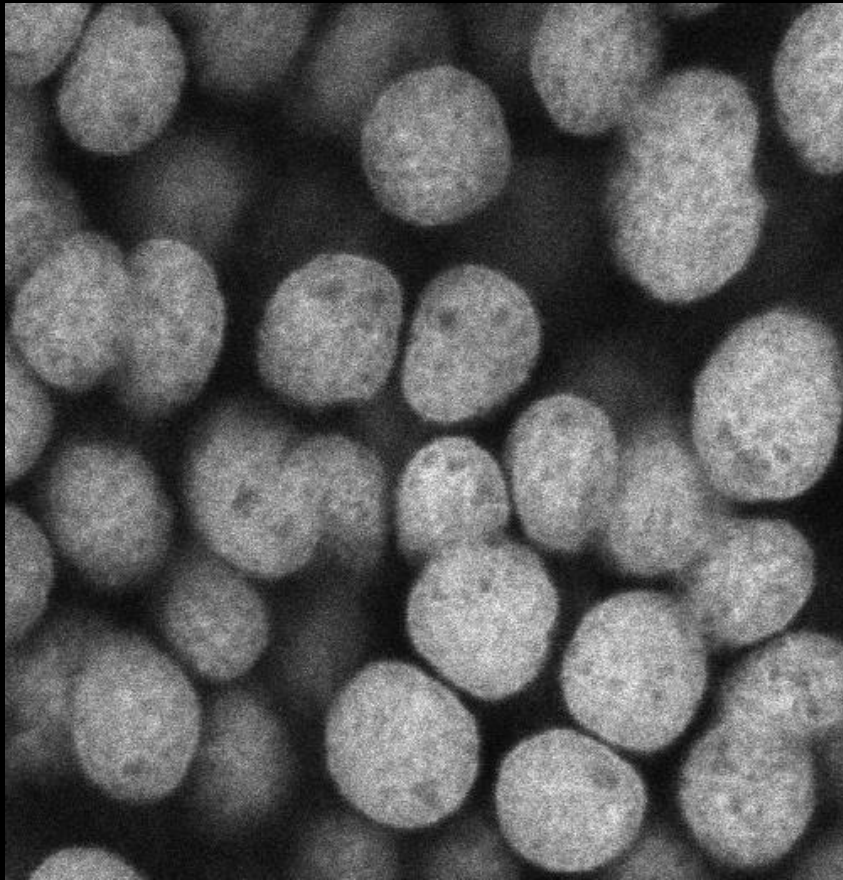


POST-PROCESSING



ANALYSIS

Pre-processing – Gaussian blur



ACQUISITION



PRE-PROCESSING



SEGMENTATION



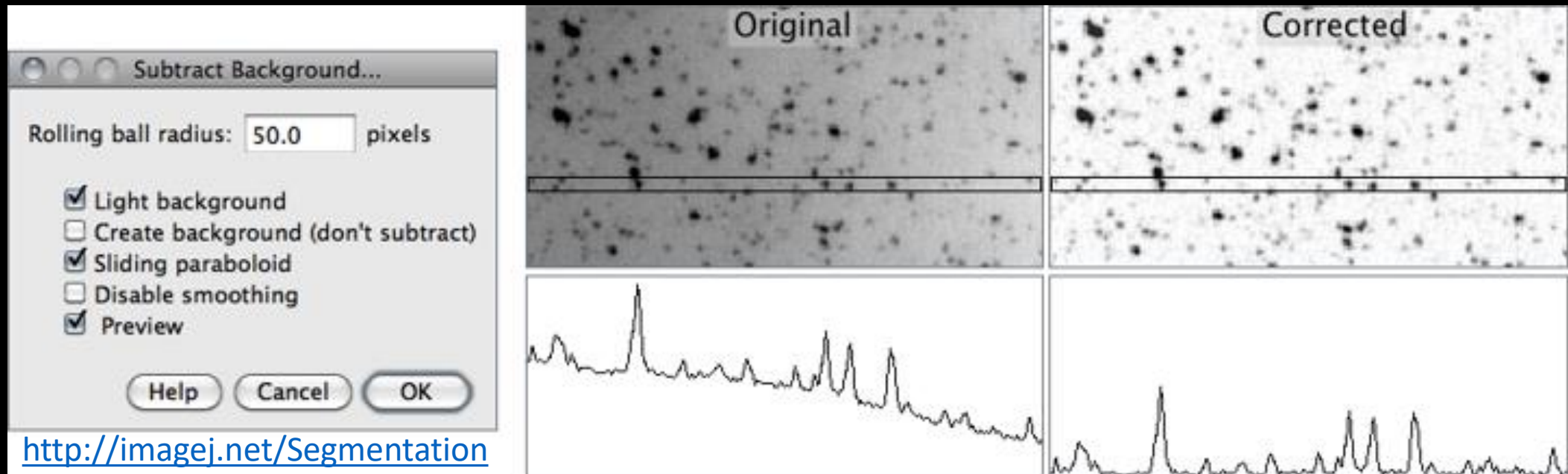
POST-PROCESSING



ANALYSIS

Pre-processing – Subtract background

- "Rolling ball" algorithm removes smooth continuous backgrounds



- In high-content analysis, background could also be calculated as an average of large number of (non-confluent) images

ACQUISITION

PRE-PROCESSING

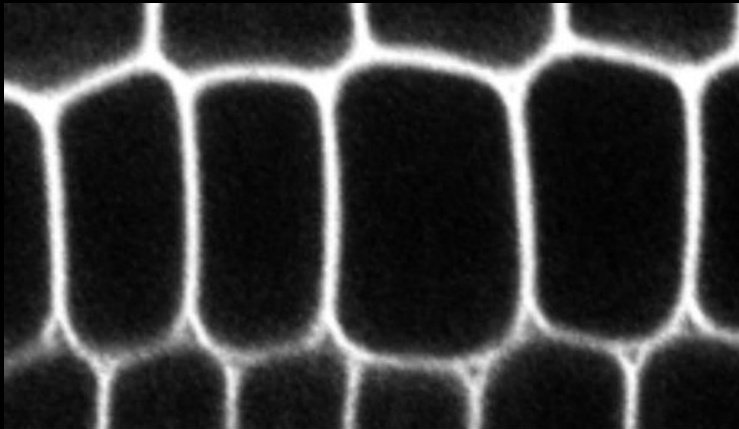
SEGMENTATION

POST-PROCESSING

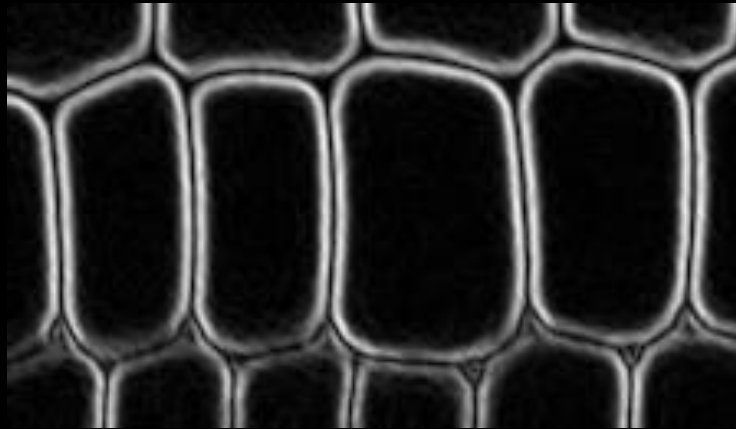
ANALYSIS

Pre-processing – Find edges

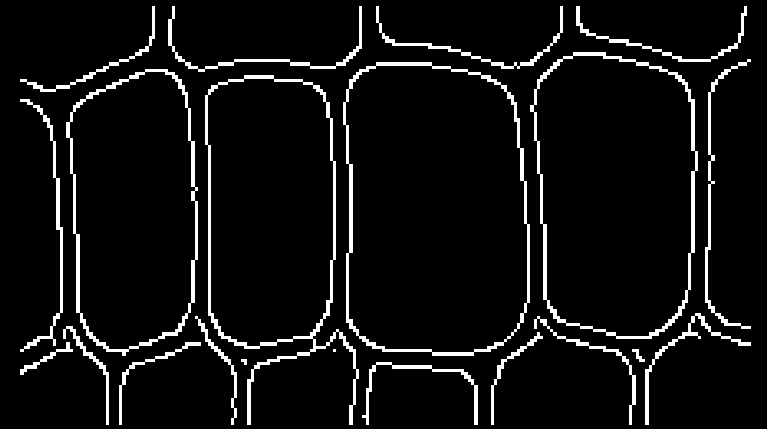
- Highlight sharp changes in intensity (by generating vertical and horizontal derivatives of the image)



Original image



Gradient image



Segmented image



Pre-processing – Deconvolution

- Error
- The
- Image



- Image restoration preserves quantitative relationships in image data
- Consider deconvolution especially if you plan to analyze colocalization

ACQUISITION



PRE-PROCESSING



SEGMENTATION



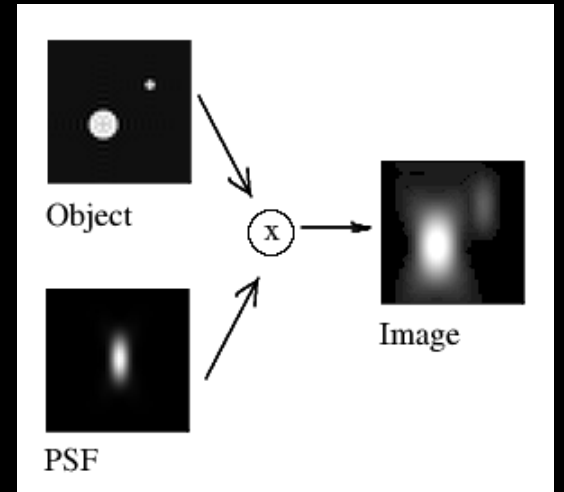
POST-PROCESSING



ANALYSIS

Pre-processing – Deconvolution

- Environmental effects and imperfections in the imaging system cause the recorded images to be degraded by blurring and noise
- The information about the blur is usually given in the form of a point spread function (PSF).
- Image deconvolution is the process of **reconstructing or estimating the true image from the degraded one**
 - Using experimental or theoretical PSF
 - Spectral filtering methods
 - Include many image **deblurring** techniques, e.g. Wiener filtering
 - Implicit assumption that the blur is spatially invariant
 - Iterative methods
 - Can be used on a much wider class of blurring models
 - Image restoration preserves quantitative relationships in image data
- **Consider deconvolution especially if you plan to analyze colocalization**



ACQUISITION

PRE-PROCESSING

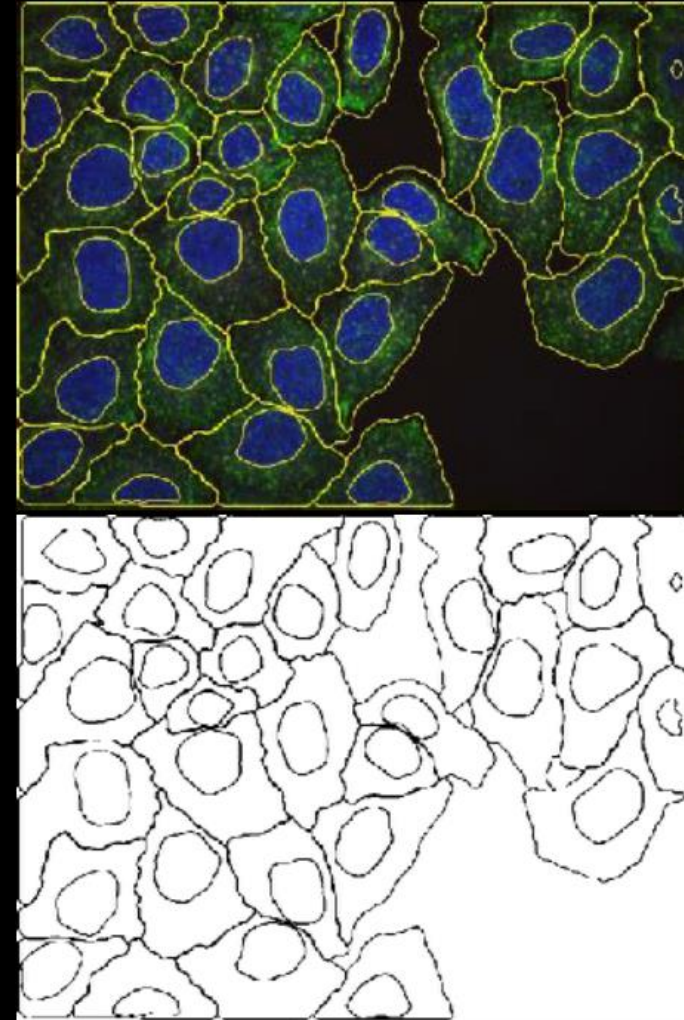
SEGMENTATION

POST-PROCESSING

ANALYSIS

Segmentation

- "Process of partitioning a digital image into multiple segments"
- Typically used to **locate objects and boundaries**



<http://imagej.net/Segmentation>

ACQUISITION



PRE-PROCESSING



SEGMENTATION



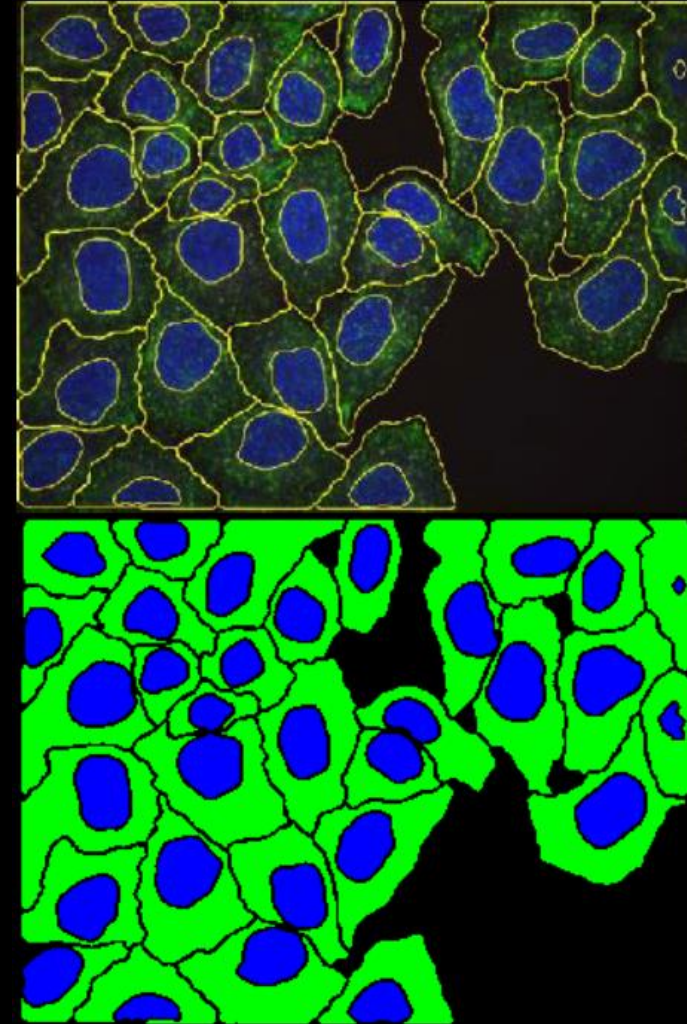
POST-PROCESSING



ANALYSIS

Segmentation

- "Process of partitioning a digital image into multiple segments"
- Typically used to **locate objects and boundaries**
- More precisely: process of **assigning a label** to every pixel in an image such that pixels with the same label share certain visual characteristics (intensity, color, texture...)



<http://imagej.net/Segmentation>

ACQUISITION



PRE-PROCESSING



SEGMENTATION



POST-PROCESSING



ANALYSIS

Segmentation

Commonly used basic methods

- Intensity thresholding (manual/automatic, global/local)
- Morphological methods (watershed)
- Shape detection (e.g. circular objects, straight lines)

More advanced methods

- Region detection using similarity criteria
- Iterative clustering methods
- Machine learning



ACQUISITION

PRE-PROCESSING

SEGMENTATION

POST-PROCESSING

ANALYSIS

Segmentation – Thresholding

- Most frequently used segmentation method is based on histogram analysis

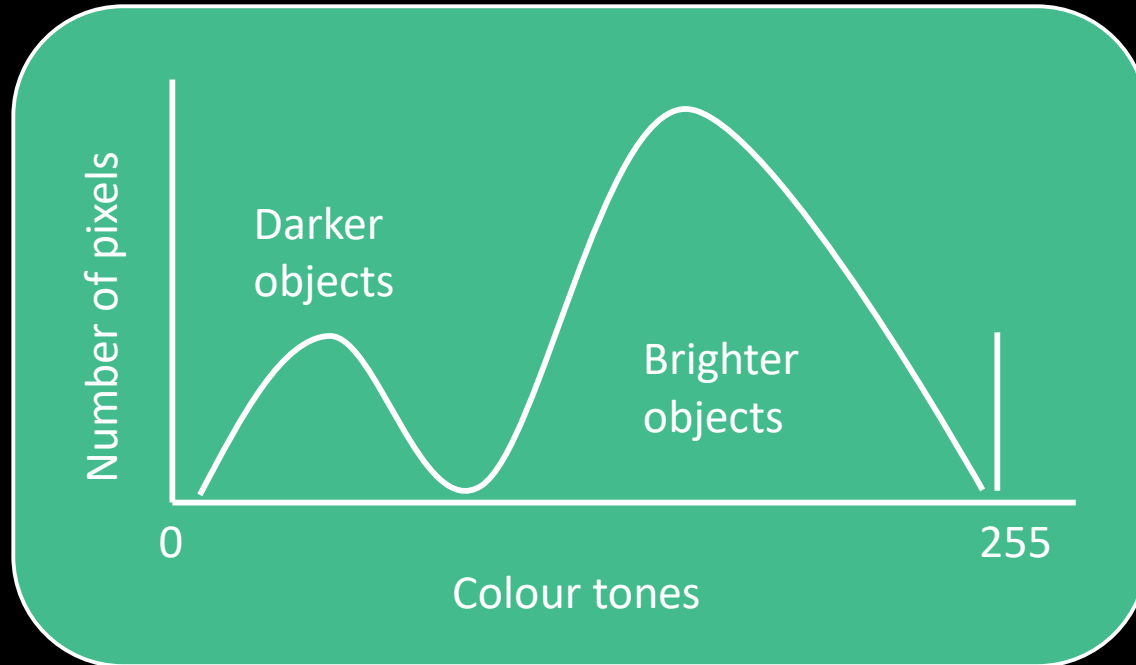


Image histogram

- Plots the number of pixels for each tonal value
- In an 8-bit image zero represents black and 255 represents white pixel value
- Peak at maximum value means signal saturation (overexposure)

ACQUISITION



PRE-PROCESSING



SEGMENTATION



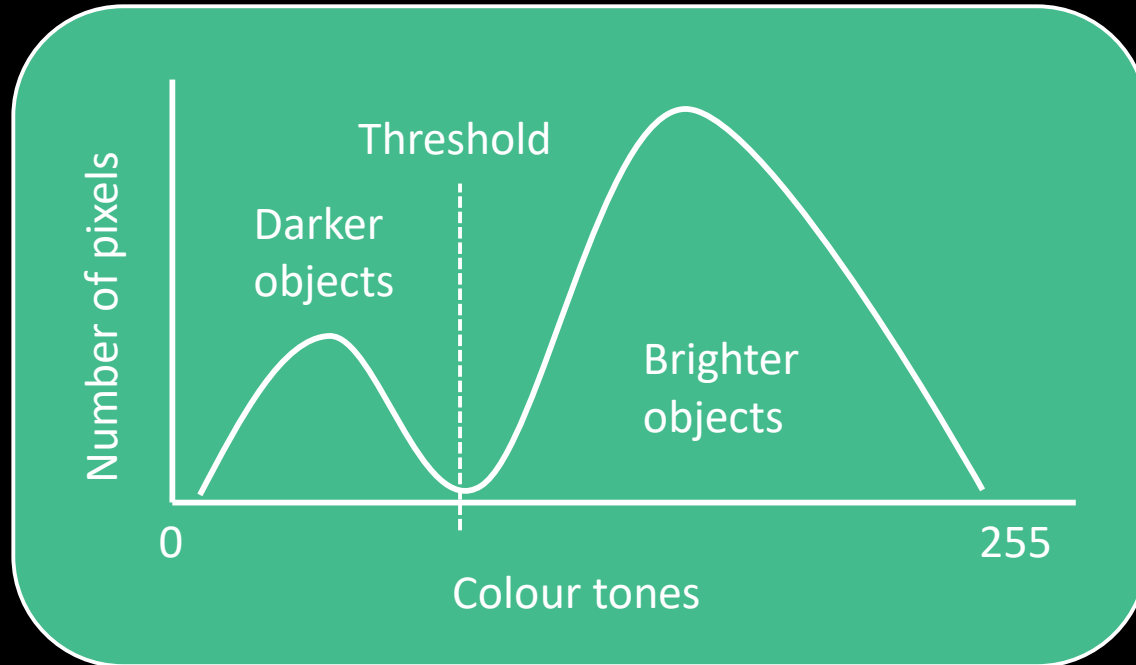
POST-PROCESSING



ANALYSIS

Segmentation – Thresholding

- Set an intensity value which separates the background and foreground



Difficulties

1. Valley may be invisible or so broad that it is difficult to locate a minimum
2. Number of minima due to the type of details in the image (multi-modal histograms)
3. Uneven illumination
4. Noise

ACQUISITION

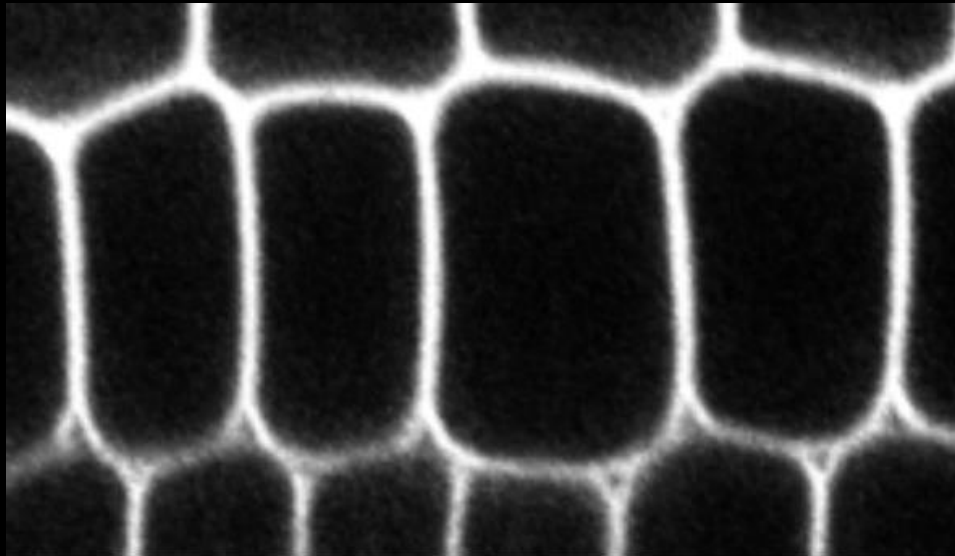
PRE-PROCESSING

SEGMENTATION

POST-PROCESSING

ANALYSIS

Segmentation – Thresholding



Original image



Thresholded image



Segmentation – Thresholding



Threshold too low



Threshold too high

ACQUISITION



PRE-PROCESSING



SEGMENTATION

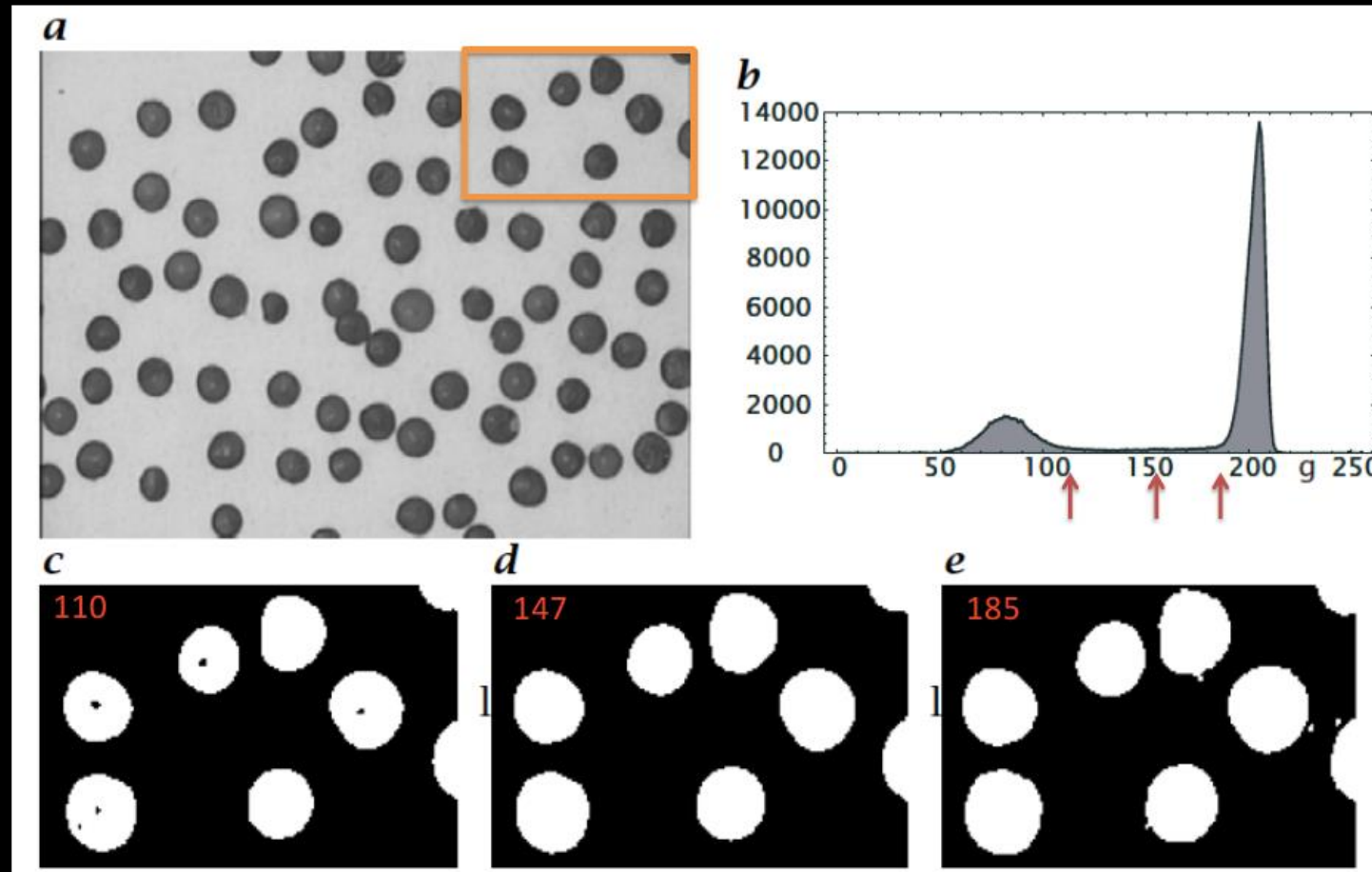


POST-PROCESSING



ANALYSIS

Segmentation – Thresholding



ACQUISITION

PRE-PROCESSING

SEGMENTATION

POST-PROCESSING

ANALYSIS

Segmentation – Global or local threshold

- Image binarization using a global or local intensity threshold?
- One **global threshold** p for mapping every pixel (i,j) into a binary (black or white) pixel, e.g. in case of an 8-bit image

$$O(i,j) = \begin{cases} 0 & \text{if } I(i,j) \leq p \\ 255 & \text{if } I(i,j) > p \end{cases}$$

- A **global threshold** can typically be used when creating large connected regions and reducing the number of small-sized darker regions (artifacts).
- A **local threshold** adapts the threshold value on each pixel (i,j) to the local image characteristics (neighboring pixels). It can be used for correcting issues like uneven illumination.

ACQUISITION



PRE-PROCESSING



SEGMENTATION



POST-PROCESSING



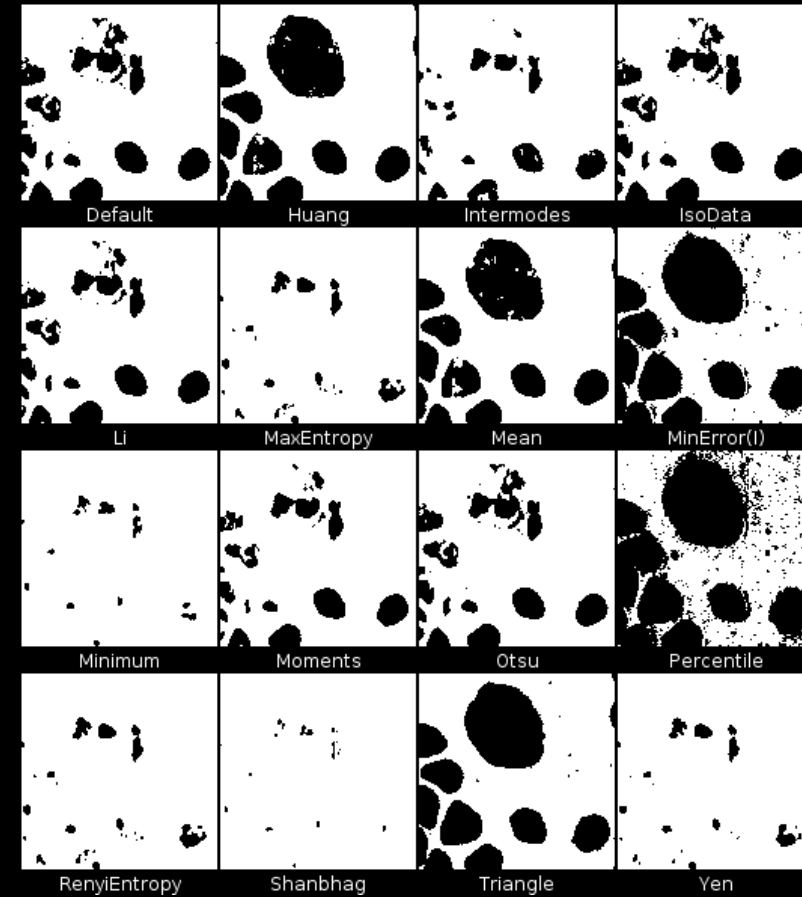
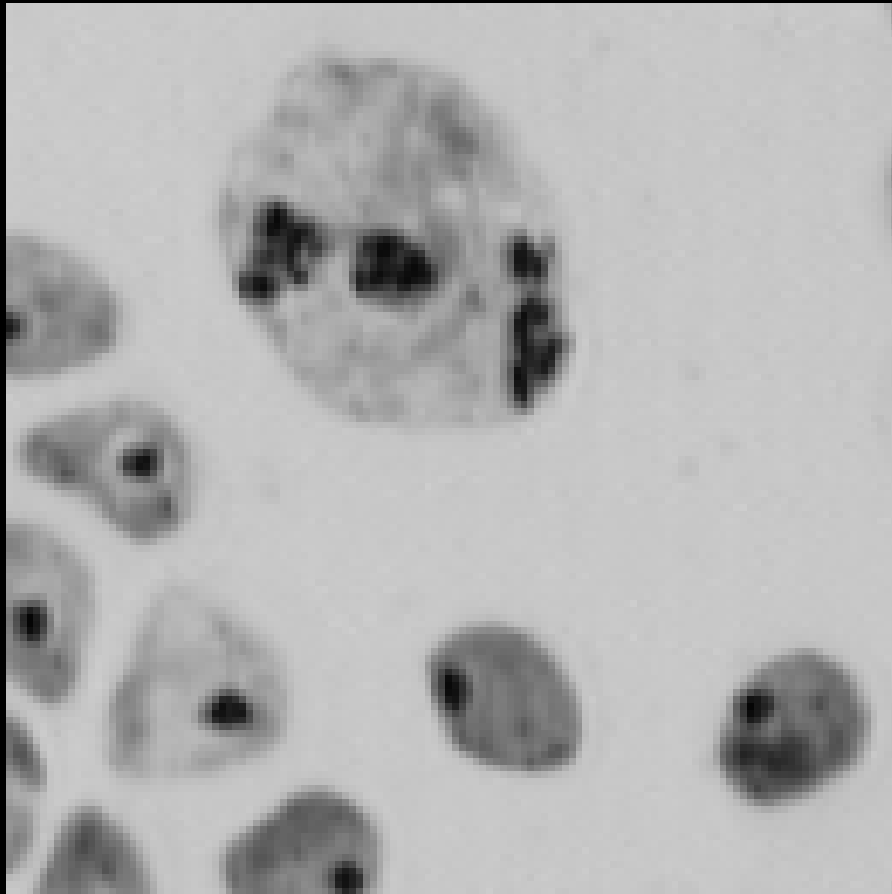
ANALYSIS

Segmentation – Automatic thresholding

- Same manual threshold over a collection of images?
 - **NOT recommended** due to fluctuations in intensity across images
- Automatic threshold by optimizing some objective criterion that can be:
 - Statistical (e.g. maximization of inter-class variance, entropy...)
 - Probabilistic (e.g. minimization of pixel classification error...)
 - Structural (e.g. circularity of detected objects...)
- How do I know whether my threshold is correct? **YOU DON'T!**
- How to choose in Fiji? **Try them ALL!**

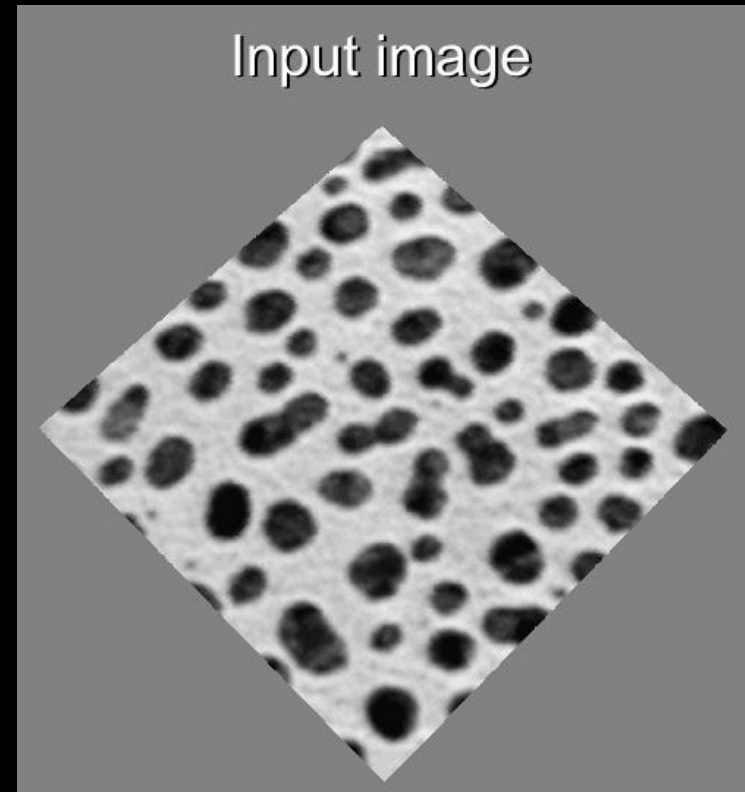


Segmentation – Automatic thresholding



Segmentation – Morphological methods

- Watershed segmentation
 - Process → Binary → Watershed
- GUI in Fiji
 - Plugins → MorphoLibJ → Segmentation → Morphological Segmentation
- Algorithm is expecting an image where the boundaries of objects present high intensity values (usually as a result of pre-processing with a gradient or edge detection filter or a distance map).



http://imagej.net/Morphological_Segmentation

ACQUISITION



PRE-PROCESSING



SEGMENTATION



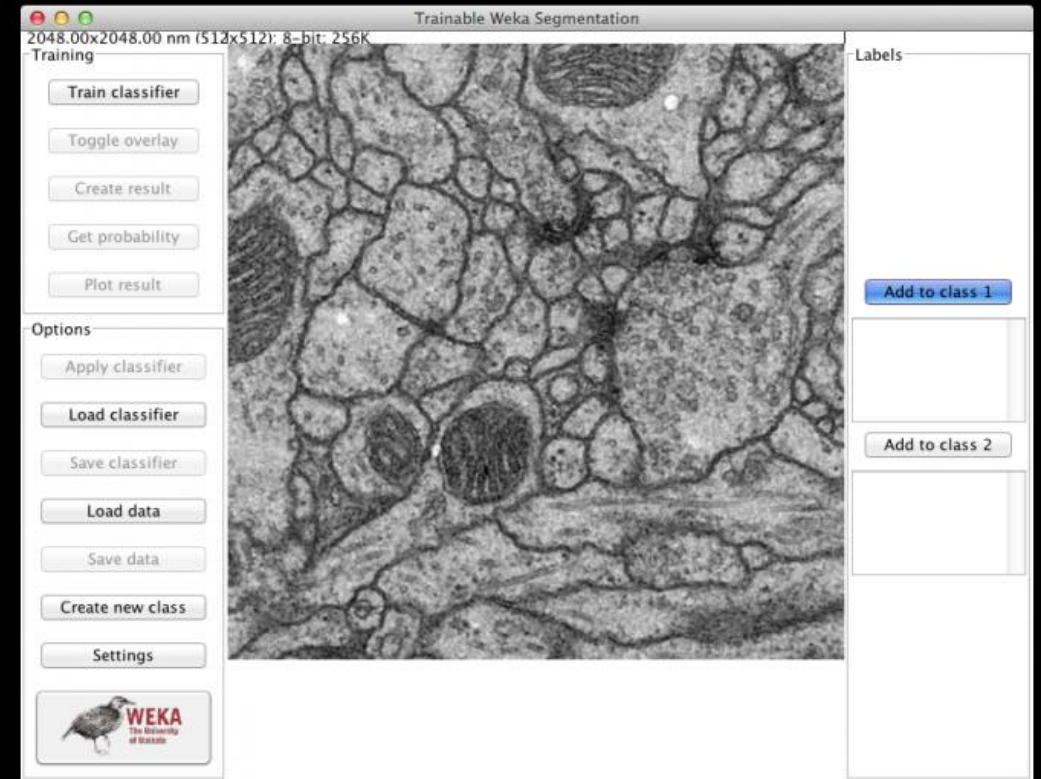
POST-PROCESSING



ANALYSIS

Segmentation – Machine learning methods

- Trainable Weka Segmentation
 - Image segmentation based on pixel classification
 - Combines a collection of machine learning algorithms with a set of selected image features to produce pixel-based segmentations
- GUI in Fiji
 - Plugins → Segmentation → Trainable Weka Segmentation



https://imagej.net/Trainable_Weka_Segmentation

ACQUISITION



PRE-PROCESSING



SEGMENTATION



POST-PROCESSING



ANALYSIS

Post-processing

Morphological reconstruction

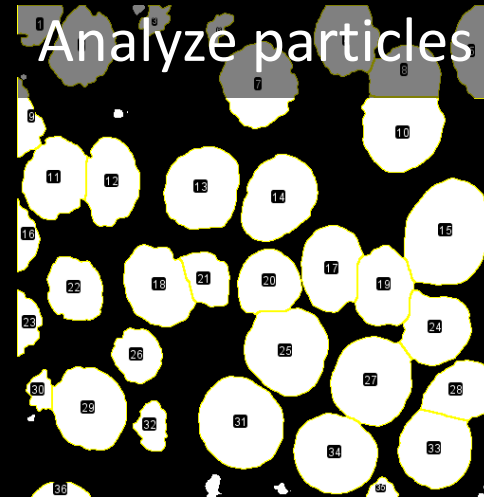
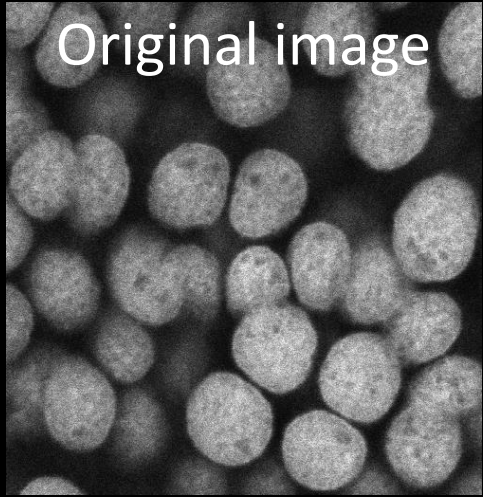
- Exclude objects on image edges
- Exclude objects based on other attributes
- Fill holes
- Separate touching objects (e.g. watershed, erosion)
- Merge separated parts (e.g. dilation)
- Regularize object shape



Analysis

- Calculations within and between selected regions of interest (ROIs)
- Object properties
 - Area, min/max/mean intensity, standard deviation, roundness etc.
 - In Fiji
 - Analyze → Set Measurements
 - Analyze → Measure
- Colocalization
 - More about that tomorrow





ACQUISITION



PRE-PROCESSING



SEGMENTATION



POST-PROCESSING



ANALYSIS

FIJI hands-on workshops

Group A: Monday 7.3. at 13.30-17.00

Group B: Wednesday 9.3. at 9.00-12.30

- Practical 1: Basics of FIJI/ImageJ (~45 min)
- Practical 2: Cell counting and characterization (~45 min)
- Practical 3: Simple macros (~60 min)

Acknowledgements

- Neubias: <http://eubias.org/NEUBIAS/>

